Comparing metagenomica and total RNA-Seq for freshwater microbiome assessments

Intro

1. Freshwater biomonitoring why
2. Entire community inclusion advocated
3. Not possible with traditional methods, but DNA based
4. Shotgun seqiencog based approaches, such as metagenomcis and metatranscriptomics, present such an option while avoiding primer bias common in metabracodng
5. metagenomics just small percentage of most common microbial barcodes ssu and lsu
6. In comparison, total RNA-Seq natural enrichment for ssu and lsu, and can be even further used for mRNA analysis (used in soil and cow gut studies)
7. Comparsion of both methods needed

(extraction of ssu reads from data, metaxa, but assembly preferred, quince 2017 and anders paper)

Introduction

Freshwater is valuable for economic productivity, ecosystem resilience, and for maintaining ecosystem services, including the supply of clean consumable water (reviewed in Dudgeon et al. 2006). However, freshwater systems are heavily impacted by agricultural and industrial pollution, habitat fragmentation (wetland drainage, river straightening, and dam building), and the introduction of invasive species (Dextrase and Mandrak, 2006; Jensen et al., 2006). Consequently, freshwater systems belong to the most threatened ecosystems with some of the highest rates of species loss (Malmqvist and Rundle, 2002).

To prevent this species loss, the natural status of freshwater bodies needs to be protected, preserved, and restored. Therefore, the ecological status of individual freshwater bodies must be identified. This knowledge help to determine if a freshwater body’s natural status is threatened and if restoration strategies need to be applied. The determination of an ecological status is accomplished by freshwater biomonitoring.

Traditionally, freshwater biomonitoring is assessed with bioindicators, most commonly benthic macroinvertebrates, but also diatoms, macrophytes, zooplankton, and fish (Bellinger and Sigee, 2015; Haury et al., 2006; Jeppesen et al., 2011; Karr, 1981; Resh and Unzicker, 1975). However, single-cellular eukaryotes (hereafter referred to as protists) and prokaryotes were proposed as nearly equally good if not better bioindicators, because they respond faster to environmental changes (Ferdous, 2009; Foissner and Berger, 1996; McArthur, 2001; Payne, 2013; Smith et al., 2015; Stoeck et al., 2018). Consequently, recent studies advocate for the inclusion of more comprehensive communities, i.e. entire microbiomes, into ecological assessments and biodiversity studies in general (Pawlowski et al., 2016; Ritter et al., 2019).

However, morphologic identification of the microbiome most protist groups and prokaryotes is not practicable because of the lack of diagnostical traits, and DNA-based methods can sometimes be the only option to distinguish organisms (Pawlowski et al., 2012; Will and Rubinoff, 2004). DNA-based biomonitoring methods have been proposed as a GOOD ADDITION to morphologic identification. While DNA Metabarcoding (Taberlet et al., 2012) has established as an effective, DNA-based biomonitoring METHOD, the method is also known to BRING ERRORS. The main issue of DNA Metabarcoding is primer bias introduced during DNA amplification, due to varying primer binding affinities (Alberdi et al. 2018; Elbrecht and Leese 2015; Krehenwinkel et al. 2017; Piñol et al. 2014; Piper et al. 2019). Additionally, PCR bias is introduced during the PCR process by GC preference of the polymerase (Nichols et al. 2018; Sze and Schloss 2019), PCR inhibition (Jane et al. 2015; Taberlet et al. 1996), and chimera formation (Piper et al. 2019; Sze and Schloss 2019). Consequently, DNA metabarcoding is unsuitable for biodiversity surveys assessing entire communities, such as microbiomes.

Shotgun sequencing-based approaches, such as metagenomics and metatranscriptomics (RNA-Seq), present an alternative to DNA Metabarcoding. These approaches enable the generation and analysis of metagenomes and metatranscriptomes of entire communities while avoiding primer and PCR bias.

Issue of these approaches: dependant on sequencing depth, and not everything can be sequenced. Especially problematic when looking at genes, barcode genes for protists and prokaryotes only make up a tiny fraction of entire sequencing set:

Standard barcode markers represent only a small portion of the read pool (0.05% for 16S ribosomal RNA (rRNA) genes (Logares et al. 2014); 0.3% for 23S/28S rRNA genes, and 1.4% for 16S/18S rRNA genes (Yilmaz et al. 2011)). The retrieval of standard barcode markers of all organisms in a sample depends on the coverage and number of reads per genetic region, which are determined by the sequencing depth, i.e. the sequenced portion of a sample. This pitfall can, however, be overcome with higher sequencing depths, which are increasingly getting cheaper.

Alternative: In comparison, total RNA-Seq natural enrichment for ssu and lsu, >85% of total RNA consists of ribosomal RNA (rRNA, component of ribosomes) (Maaløe 1979) and can be even further used for mRNA analysis (used in soil and cow gut studies). Also active part of community: Total-RNA sequencing can be used to identify the active part of a community (Gomez-Silvan et al. 2018), based on the fact that the abundance of rRNA in a cell (as in the number of ribosomes) is an indicator for cell growth and activity (Milo and Phillips 2015). This signal contrasts with that from metagenomics, which targets the present community, including dead and/or inactive cells and extracellular DNA, which can make up high portions of microbial DNA datasets (up to >90%; Carini et al. 2016; Torti, Lever, and Jørgensen 2015).

Comparsion of both methods needed for freshwater microbiome assessments. Here we compare both using mock and real freshwater microbiomes, testing different pipelines. Many pipelines have been compared for metagenomics REF, but not for total RNA Seq

Diskussion:

Metabarcoding for entore communities with multi-marker approaches

Rna hard to handle